

# *Potential landscape-scale pollinator networks across Great Britain: structure, stability and influence of agricultural land cover*

Article

Published Version

Creative Commons: Attribution 4.0 (CC-BY)

Open Access

Redhead, J. W., Woodcock, B. A., Pocock, M. J. O., Pywell, R. F., Vanbergen, A. J. and Oliver, T. H. (2018) Potential landscape-scale pollinator networks across Great Britain: structure, stability and influence of agricultural land cover. *Ecology Letters*, 21 (12). pp. 1821-1832. ISSN 1461-0248 doi: <https://doi.org/10.1111/ele.13157> Available at <https://centaur.reading.ac.uk/78923/>

It is advisable to refer to the publisher's version if you intend to cite from the work. See [Guidance on citing](#).

To link to this article DOI: <http://dx.doi.org/10.1111/ele.13157>

Publisher: Wiley

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

[www.reading.ac.uk/centaur](http://www.reading.ac.uk/centaur)

## **CentAUR**

Central Archive at the University of Reading

Reading's research outputs online

## LETTER

# Potential landscape-scale pollinator networks across Great Britain: structure, stability and influence of agricultural land cover

John W. Redhead,<sup>1,4\*</sup> Ben A. Woodcock,<sup>1</sup> Michael J.O. Pocock,<sup>1</sup> Richard F. Pywell,<sup>1</sup> Adam J. Vanbergen<sup>2,3</sup> and Tom H. Oliver<sup>1,4</sup>

### Abstract

Understanding spatial variation in the structure and stability of plant–pollinator networks, and their relationship with anthropogenic drivers, is key for maintaining pollination services and mitigating declines. Constructing sufficient networks to examine patterns over large spatial scales remains challenging. Using biological records (citizen science), we constructed potential plant–pollinator networks at 10 km resolution across Great Britain, comprising all potential interactions inferred from recorded floral visitation and species co-occurrence. We calculated network metrics (species richness, connectance, pollinator and plant generality) and adapted existing methods to assess robustness to sequences of simulated plant extinctions across multiple networks. We found positive relationships between agricultural land cover and both pollinator generality and robustness to extinctions under several extinction scenarios. Increased robustness was attributable to changes in plant community composition (fewer extinction-prone species) and network structure (increased pollinator generality). Thus, traits enabling persistence in highly agricultural landscapes can confer robustness to potential future perturbations on plant–pollinator networks.

### Keywords

Citizen science, ecological networks, ecosystem services, food webs, pollination, resilience, stability.

*Ecology Letters* (2018)

## INTRODUCTION

Insect pollinators face many threats that may jeopardise the crucial ecosystem service they provide to crops and wild plants (Vanbergen *et al.* 2013; Gill *et al.* 2016; Potts *et al.* 2016). The stability of pollinator communities and the service they deliver is mediated by the structure of ecological networks formed by interactions between pollinator and plant species (Vázquez *et al.* 2009; Vanbergen *et al.* 2017). Understanding such networks is important to predict the risks associated with threats to pollinators (Gill *et al.* 2016). Analysis of plant–pollinator networks has provided insights into their structure and potential stability under actual or simulated environmental change, including extinctions (e.g. Memmott *et al.* 2004; Kaiser-Bunbury *et al.* 2010), climate change (e.g. Memmott *et al.* 2007), habitat change (e.g. Forup *et al.* 2008; Vanbergen *et al.* 2017) and restoration (Kaiser-Bunbury *et al.* 2017).

Studies traditionally rely on obtaining well-characterised networks from field surveys, which are time consuming and costly to construct (Vázquez *et al.* 2009; Burkle & Alarcón 2011). Constructing networks replicated across larger spatial scales remains a daunting prospect (Burkle & Alarcón 2011), particularly at the regional and national scales relevant to

land use and conservation policy-making. Although broad geographical patterns in plant–pollinator network properties have been identified across biomes (Olesen & Jordano 2002; Welti & Joern 2015) or within landscapes (Burkle & Alarcón 2011; Carstensen *et al.* 2014; Trøjelsgaard *et al.* 2015; Kaiser-Bunbury *et al.* 2017), these still rely on a comparatively limited number of empirical plant–pollinator networks.

Of particular interest in understanding spatial variability in plant–pollinator networks is the contrast between the benefits of insect pollinators to agricultural crops (Kremen *et al.* 2002; Winfree 2008; Eilers *et al.* 2011) and the negative impacts of intensive agriculture on pollinators (Kluser & Peduzzi 2007; Potts *et al.* 2010; Gill *et al.* 2016). However, we have very little knowledge of how plant–pollinator networks are affected by agriculture at landscape scales (e.g. > 1 km<sup>2</sup>) or whether networks comprising species that pollinate agricultural crops are representative of the wider pollinator community (Kleijn *et al.* 2015; Gill *et al.* 2016).

Lack of information on ecological interactions across larger spatial, temporal and taxonomic scales, termed the ‘Eltonian shortfall’, represent key gaps in our large-scale knowledge of biodiversity (Hortal *et al.* 2015). Moreover, there are limitations on the extent to which different data sources can be combined to analyse multiple networks because data

<sup>1</sup>NERC Centre for Ecology and Hydrology, Maclean Building, Wallingford, Oxfordshire OX108BB, UK

<sup>2</sup>NERC Centre for Ecology and Hydrology, Bush Estate, Penicuik, Midlothian EH26 0QB, UK

<sup>3</sup>Agroécologie, AgroSup Dijon, INRA, Univ. Bourgogne Franche-Comté, F-21000 Dijon, France

<sup>4</sup>School of Biological Sciences, University of Reading, Harborne Building, Reading, Berkshire RG6 6AS, UK

\*Correspondence: Email: johdhe@ceh.ac.uk

collection methods can introduce potential biases (Hortal *et al.* 2015). While there are exciting possibilities for molecular techniques to increase the speed and accuracy with which plant–pollinator networks can be constructed (Keller *et al.* 2015; Richardson *et al.* 2015; Bohan *et al.* 2017; Pornon *et al.* 2017), these are yet to be realised across larger spatial scales.

Biological records (i.e. records submitted to voluntary recording schemes, a form of ‘citizen science’) provide a valuable resource for analysing large-scale patterns in time and space (Bishop *et al.* 2013; Tulloch *et al.* 2013; Powney & Isaac 2015). Records consist of species’ identification, date and location (hereafter ‘occurrence’ data) and provide large volumes of data over a wide spatial coverage, equivalent to innumerable hours of field survey. Methods to control for variation in recorder effort and to infer ecological signals from occurrence data are rapidly emerging (e.g. Isaac *et al.* 2014; Dyer *et al.* 2016), but hitherto their potential as a source of data on ecological networks is untapped (Gray *et al.* 2014).

Here, we constructed potential plant–pollinator networks for every 10 km-by-10 km grid square (‘hectad’) in Great Britain (GB) using interactions from a 30-year, long-term national data set of occurrence records of pollinating insects (bees, butterflies and hoverflies). Instead of inferring species interactions from spatial co-occurrence (Morales-Castilla *et al.* 2015; Morueta-Holme *et al.* 2016) we used metadata from records that detailed flower visitation as a proxy of pollination (Ballantyne *et al.* 2015). These networks are ‘potential’ in that we acknowledge their limitations in terms of assumptions that constrain their biological realism. However, while the structure of each potential network may be subject to errors, we aimed to minimise bias affecting comparisons across replicate networks. We used these potential networks to address three questions. First, does network structure and stability vary spatially across GB? Second, is network stability reduced by greater agricultural land cover, a major driver of plant and pollinator declines (Kluser & Peduzzi 2007; Potts *et al.* 2010; Vanbergen *et al.* 2013; Ollerton *et al.* 2014)? Finally, are the structure and stability of networks comprising crop–pollinator species consistent with those of the wider pollinator community?

## MATERIALS AND METHODS

### Constructing a plant–pollinator interactions database

We constructed a national-scale (GB) plant–pollinator interaction database defining which species of pollinator visit which species of plant (Fig. 1). Data were mostly (73%) sourced from biological records. Specifically, these were species observations submitted to the Bees, Wasps and Ants Recording Society (BWARS), Butterflies for the New Millennium (BNM, Asher 1997) and Hoverfly Recording Scheme (HRS), with plant interactions recorded as incidental metadata. Interactions were inferred by algorithmically screening metadata for valid scientific or vernacular plant names (or widely used synonyms or abbreviations thereof), followed by data cleaning (see Appendix S1). Remaining interaction data were obtained from books (e.g. Morris 1998), papers (e.g. Carvell 2002) and unpublished experimental data. Where interactions were

recorded only to plant genus we assumed, given the rarity of pollinators specialised to the level of individual plant species (Waser *et al.* 1996; Minckley & Roulston 2006), that these were indicative of interactions with all plant species within the genus that were present in the data set (full details in Appendix S1). These inferred interactions comprised 6487 unique interactions (39%) within the full data set.

Our final plant–pollinator interactions database contained 16,712 unique interactions, involving 485 pollinator species (206 bees, 56 butterflies and 223 hoverflies) and 499 plant species. This total comprises approximately 76, 92, 81 and 55% of GB bee, butterfly, hoverfly and insect-pollinated plant species respectively (Fitter & Peat 1994; Stubbs & Falk 2002; Thomas 2010; Falk 2015). We explored the completeness of our interactions database by calculating interaction accumulation curves across all records used to construct the database (i.e. pollinator occurrences where we were able to identify a valid plant interaction) and for each plant and pollinator species separately (Appendix S2). Results suggested that our database captured around 60% of estimated total interactions (mean 62% for pollinators, 57% for plants), comparable to studies which performed high-effort, multitemporal field sampling of individual networks (Chacoff *et al.* 2012; Falcão *et al.* 2016).

### Modelling plant and pollinator occurrence

For all species in the interactions database, we obtained occurrence data from BWARS, HRS, BNM and, for plants, the Botanical Society of Britain and Ireland (Fig. 1). Data records were restricted to 1985 onwards, covering the vast majority of records while excluding occurrences of species that may have been more widespread prior to major changes in GB land use (Robinson & Sutherland 2002; Ollerton *et al.* 2014). Occurrence data were modelled to account for spatial bias in recorder effort using the FRESALO algorithms (Hill 2012), implemented in the SPARTA (v0.1.30 August *et al.* 2015b) package of R (v3.4.0 R Core Team 2017). FRESALO weights by recorder effort to estimate trends and probability of occurrence in under-recorded areas (for validation of FRESALO for different groups and through simulation see Hill 2012; Fox *et al.* 2014; Isaac *et al.* 2014; Dyer *et al.* 2016). We used the CEH Land Cover Map (LCM2007, Morton *et al.* 2011) as input data for FRESALO’s calculation of neighbourhoods of ecologically similar hectads (see August *et al.* 2015b; Dyer *et al.* 2016). For each species, FRESALO produces a probability of occurrence per hectad. To transform this to presence/absence, we assigned a species as present in a hectad if its probability of occurrence was greater than a set threshold (see Appendix S3).

### Constructing potential networks

We used lists of modelled plant and pollinator species presence per hectad to filter the interactions database (as derived from plant associations in biological records) and create a potential plant–pollinator network for each hectad (Fig. 1). Networks were unweighted (i.e. interaction matrices consisting of ones and zeros), this being the most conservative

interpretation of our interaction data because the frequency with which an interaction was recorded is unlikely to provide reliable quantitative information on abundance, due to differences in detectability, recorder bias and data sources.

### Network structure metrics

From the networks constructed in each of 2823 GB hectads, we used the R package *bipartite* (v2.07, Dormann *et al.* 2009) to calculate the following metrics (Bersier *et al.* 2002; Dunne *et al.* 2002; Tylianakis *et al.* 2007):

- (1) Species richness: total number of plant and pollinator species in the network
- (2) Connectance: proportion of possible links which are realised
- (3) Pollinator generality: mean number of plants per pollinator
- (4) Plant generality: mean number of pollinators per plant

While other, more complex metrics of network structure (e.g. nestedness, modularity) have been implicated in stability they can be comparatively insensitive to spatial or temporal change (Kaartinen & Roslin 2012; Morris *et al.* 2014; Kemp *et al.* 2017). Preliminary analyses confirmed that nestedness and modularity showed little variation even for networks greatly differing in the metrics listed above.

### Robustness to simulated extinctions

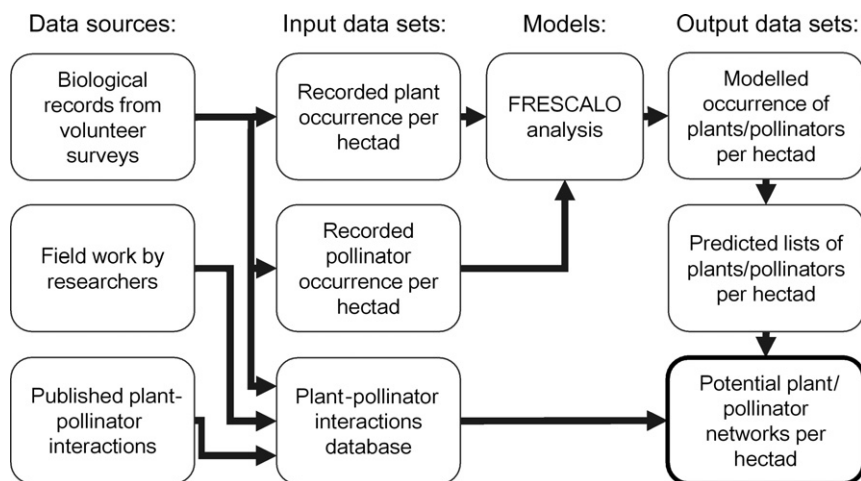
To derive a metric of network stability, we assessed the impact of simulated extinctions of plants. This was a measure similar to robustness (following Memmott *et al.* 2004; Burgos *et al.* 2007), but differing in that sequential simulated extinctions were ordered according to a complete ‘global’ list of plants (i.e. across all hectads) and not just those in each ‘local’ network (i.e. individual hectad) (Fig. 2). This approach meant the same extinction scenario was universally applied across hectads and enables comparisons across networks with different plant communities. We term this approach and the

resulting metric ‘robustness to global simulated extinctions’ ( $R_g$  from here on) to avoid confusion with the usual approach. For comparison, we also calculated ‘local’ robustness ( $R_l$  from here on) following Memmott *et al.* (2004) with randomised extinction of plants within each hectad. We focussed on simulating plant extinctions because many of the major impacts of agriculture indirectly affect pollinators via altered plant communities (Potts *et al.* 2010; Vanbergen *et al.* 2013, 2017), as would restoration of agricultural plant–pollinator networks in practice (Kremen *et al.* 2002; Forup *et al.* 2008; Menz *et al.* 2011; Kaiser-Bunbury *et al.* 2017).

As well as randomised plant extinctions from the global list ( $R_g R$ ), we conducted simulated extinctions by ordering the complete list of 499 plant species according to potential predictors of future plant declines under three scenarios:

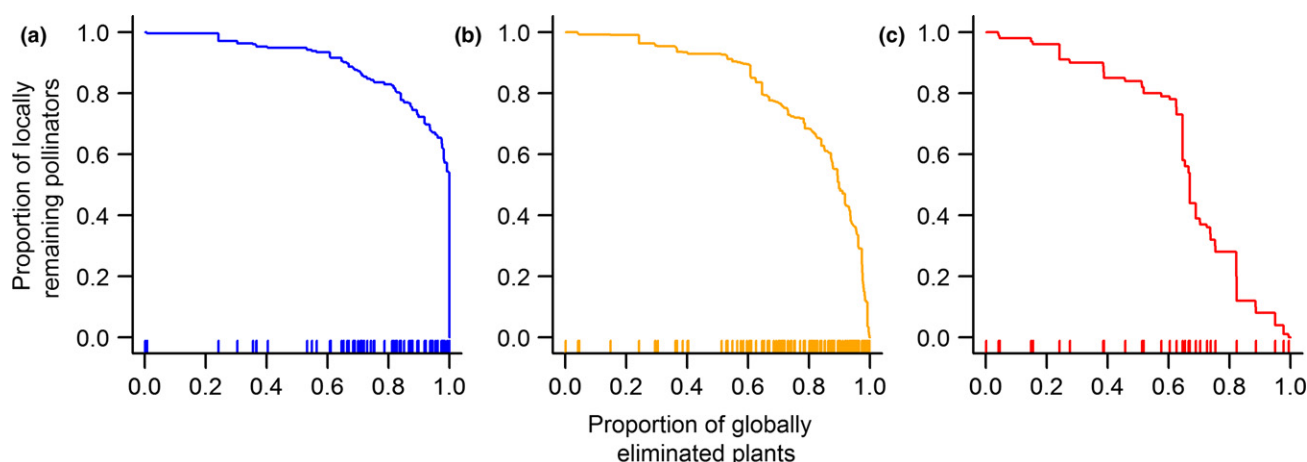
- (1) historic distribution trend (1985–2015) estimated using FRESALO (Hill 2012; Isaac *et al.* 2014), extinctions occurring first for plants showing the greatest historic decline ( $R_g Trend$ );
- (2) soil fertility tolerance based on Ellenberg N values (Hill *et al.* 2004), extinctions occurring first for plants preferring low soil fertility, as historically observed in GB flora (Stevens *et al.* 2006; Maskell *et al.* 2010) ( $R_g N$ );
- (3) drought tolerance based on Ellenberg F values (Hill *et al.* 2004), extinctions occurring first for plants preferring moister conditions expected to suffer under climate change (Thuiller *et al.* 2005; Watts *et al.* 2015) ( $R_g F$ ).

For each hectad-level network, plants were sequentially extirpated from the global list in the order determined by each scenario. After each plant extinction, any pollinator species with no remaining links were removed from the network. We assessed  $R_g$  as the area under the curve (Burgos *et al.* 2007) of pollinators remaining in the local network against plants removed from the global sequence (Fig. 2). This process was repeated 100 times per scenario, with random ordering of plant species with tied trends or Ellenberg values (or of the entire list for randomised plant extinctions;  $R_g R$ ).



**Figure 1** Schematic showing steps in construction of potential plant–pollinator networks for every 10 × 10 km cell (‘hectad’) in Great Britain from biological records.





**Figure 2** Example extinction curves (coloured lines) for three example hectads (a, b, c) with different plant–pollinator communities and correspondingly different robustness to global simulated extinctions as measured by area under extinction curve (a:  $R_g = 0.90$ , b:  $R_g = 0.82$ , c:  $R_g = 0.66$ ). Plant extinctions are ordered by trend (i.e. most strongly declining are eliminated first). Tick marks along the x-axes indicate where simulated plant extinction from the global list resulted in an extinction from the hectad.

Following individual plant extinctions, there is potential for pollinators to switch between plants and rewire networks (Thomsen *et al.* 2017). Other authors have used regional information to inform the likelihood of rewiring in local networks (Kaiser-Bunbury *et al.* 2010). However, our potential networks already implicitly incorporate some of this capacity for rewiring because each local network contains information from all recorded interactions across GB over three decades. Another approach is to create putative novel interactions from plant–pollinator traits but, given the assumptions and uncertainties involved, it is difficult to assess whether such rewiring scenarios are more ecologically meaningful than using only observed interactions. We conducted supplementary analyses exploring additional trait-based network rewiring scenarios but found that spatial patterns in robustness metrics were largely unaffected (Appendix S4).

Two main sources of error in our hectad-level potential networks are the methods used to model occurrence and the database used to assign interactions. Consequently, we assessed the impact of these sources of uncertainty independently and in combination by performing, for each hectad, 100 randomised resamples of species according to FRES-CALO probability of occurrence, and of interactions proportional to the number of times they were recorded (see Appendix S5).

### Crop-pollinators

We repeated our analyses by calculating hectad-level network metrics, including  $R_g$  and  $R_b$ , for potential networks consisting solely of interactions involving known crop-pollinators. This allowed us to explore whether the structure and stability of crop–pollinator networks was similar to the wider plant–pollinator networks in which they were embedded. Bees are generally considered the most important contributors to crop pollination (Free 1993; but see Rader *et al.* 2016) and their predominance in the pollination of GB crops is well supported (Woodcock *et al.* 2013; Garratt *et al.* 2014). Crop-pollinators

were determined from a published list of bee species with the highest contribution to crop production value (Kleijn *et al.* 2015) for major GB insect-pollinated crops (oilseed rape, field bean, apple and strawberry). Our interactions database included 23 such species from 5 genera (see Fig. S5 for full species list). We then compared metrics for crop–pollinator networks, overall plant–pollinator networks and bee-only networks. Because crop–pollinator networks are considerably less speciose, we resampled the bee-only network for each hectad 100 times, with a random selection of pollinators equal in number to crop-pollinator species in the hectad, and then calculated mean resampled network metrics for comparison. The number of plants in each resampled network was allowed to vary depending on interactions with the selected pollinators, as attempting to limit plants to the number in the crop-pollinator networks would severely restrict the number of resampled networks and constrain resultant network metrics.

### Statistical analysis

Network metrics were modelled independently against agricultural coverage, using linear mixed-effects models in the *nlme* R package (v3.1 Pinheiro *et al.* 2015). We derived coverage of agricultural land (arable + improved grassland) per hectad from LCM2007 and used this as a fixed effect explanatory variable, along with an optional quadratic term, which was retained in models if significant. To account for the potential influence of other environmental variables on network structure and response to agricultural coverage, we assigned each hectad to an environmental zone, using a pre-existing classification (Bunce *et al.* 2007). Environmental zone was then included as a random factor in all models, with variable slope and intercept. Some of the network metrics we used are sensitive to the size of the network (Jordano 1987; Olesen & Jordano 2002; Forup *et al.* 2008; Morris *et al.* 2014), so models were compared with and without total species richness as a fixed covariate. Environmental zones represented by < 30 hectads were considered to have insufficient sample size for

robust analysis and were excluded, as were hectads with > 50% coverage of sea, giving a final sample of 2290 hectads. All variables were standardised to mean of zero and standard deviation of one, to facilitate comparison of model coefficients. Each model was compared using a likelihood-ratio test against a model consisting only of the random effect and species richness, to determine the impact of incorporating agricultural coverage on model fit. We also applied randomisation tests (Fortin & Jacquez 2000) to account for potential complex spatial autocorrelation patterns arising from how FRES-CALO defines neighbourhoods based on spatial proximity and biological similarity (see Appendix S6).

## RESULTS

### Spatial patterns in network properties

Variation in plant and pollinator species richness conformed to known clines across GB (i.e. higher richness in the south and at lower altitudes, Fig. 3). Spatial patterns for connectance, pollinator generality and plant generality showed similar latitudinal and altitudinal trends to species richness (Fig. 3), and a significant correlation with total plant-pollinator species richness (Pearson's  $r = -0.95$ ;  $0.82$ ;  $0.97$ , respectively,  $n = 2290$ ,  $P < 0.001$  in all cases).

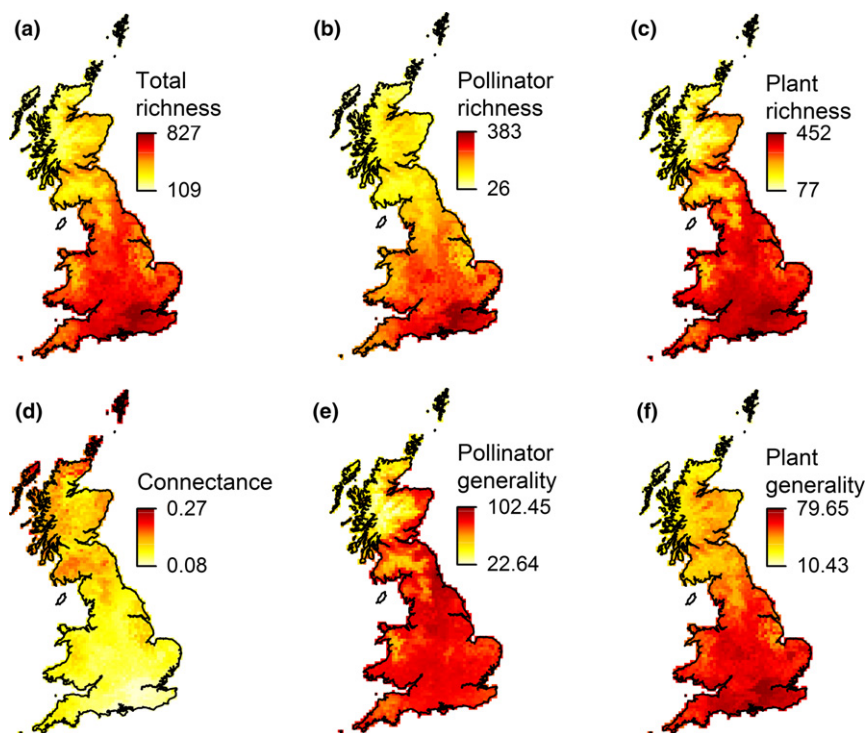
Robustness to global simulated extinctions ( $R_g$ ) also showed spatial variation across GB, with more variation than would be expected under simple conformity to species richness, latitude or altitude (Fig. 4a, c, e, g and i). The different extinction sequences gave mean  $R_g$  scores across hectads of  $0.84$ ,

$0.92$ ,  $0.85$  and  $0.93$  for extinctions ordered by trend ( $R_{g\ Trend}$ ), Ellenberg N ( $R_{g\ N}$ ), Ellenberg F ( $R_{g\ F}$ ) and randomised extinctions ( $R_{g\ R}$ ), respectively (range across hectads  $0.66$ – $0.97$  across all four  $R_g$  measures), but with varying spatial patterns (Fig. 4). Robustness to randomised local extinctions ( $R_l$ ) showed a very similar range of values and spatial patterns to  $R_{g\ R}$  (Fig. 4i).

### Effect of agricultural land cover on network properties and robustness

Pollinator generality and all five measures of robustness to simulated extinctions showed significant positive relationships with agricultural coverage (Table 1). All relationships apart from  $R_{g\ F}$  included a significant negative quadratic term (Table 1) indicating a levelling off of the relationship as agricultural coverage approaches 100% (Fig. 5a, b, c, e, f). These results suggest that pollinator communities in more highly agricultural landscapes are more generalist and that, under all our extinction scenarios, hectads with a higher coverage of agricultural land lost pollinators less quickly than other hectads in the same environmental zone. This effect appeared most pronounced for  $R_{g\ Trend}$  and  $R_{g\ N}$  (Fig. 5b, c). Neither plant generality nor connectance showed a significant relationship with proportion of agricultural coverage (Table 1).

While species richness was a significant covariate in models for all network metrics, its inclusion did not qualitatively change the relationships with agricultural coverage. For all models, likelihood ratio tests and randomisation tests



**Figure 3** Network properties per hectad across GB: a) total species richness of plants and pollinators combined, b) richness of pollinators and c) richness of plants, d) network connectance, e) pollinator generality and f) plant generality. Lighter colours indicate lower values, darker colours indicate higher, with a linear colour stretch between maximum and minimum values.

generally corroborated the significance of the individual model coefficients for agricultural coverage (Table 1). There was no significant relationship between agricultural coverage and

species richness of plants, pollinators or both combined, once environmental zones were accounted for (Table 1).

There were significant relationships between agricultural coverage and the mean values across the plant community per hectad of the trait and trend values used to order extinction sequences (Table 1). This indicates that agricultural coverage influences the relative position of the plant community in our global extinction sequences. Standard deviations in these values within hectads showed a significant, negative relationship only for Ellenberg  $N$ , suggesting hectads with higher agricultural cover not only host communities with higher average fertility tolerance, but also show significantly less variation in fertility tolerance between plant species.

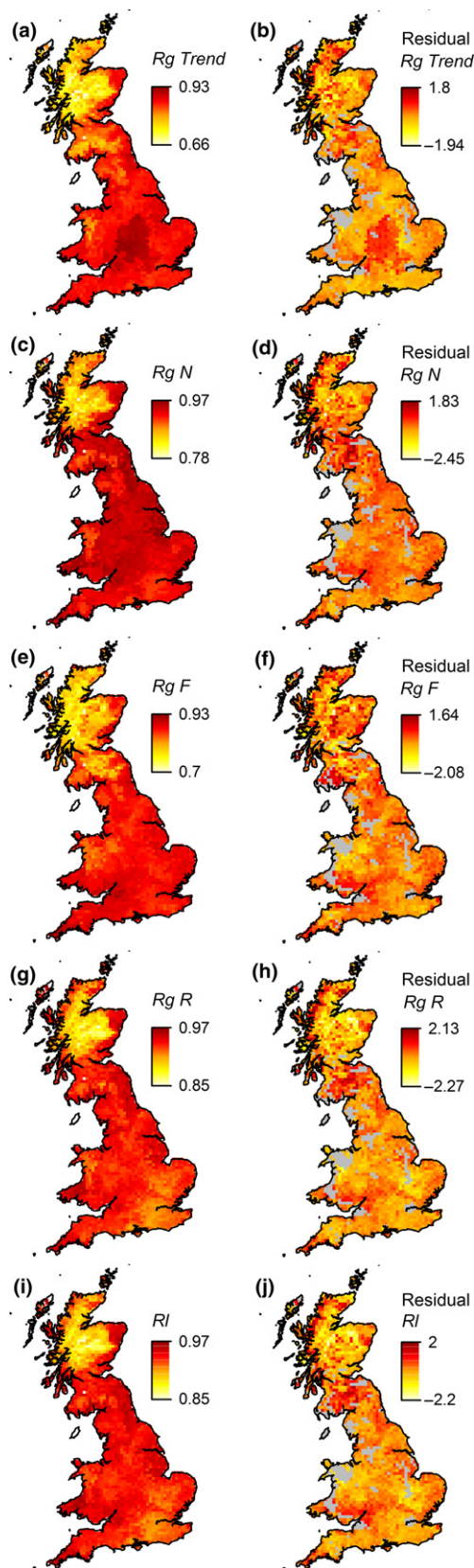
### Crop–pollinator networks

Subsets of networks consisting of only crop-pollinating bees and the plants they visit showed significant differences in their properties from complete hectad-level networks, or from randomly resampled networks of equivalent bee species richness. Crop–pollinator networks showed significantly higher plant species richness than the randomly resampled bee networks (pairwise  $t$ -test;  $t = 133.57$ ,  $P < 0.001$ , d.f. = 2750), as well as higher connectance and pollinator and plant generalities (pairwise  $t$ -test;  $t = 159.54$ ,  $155.48$  and  $116.35$  for connectance, pollinator generality and plant generality, respectively; d.f. = 2750 and  $P < 0.001$  in all cases). Robustness values for crop–pollinator networks were significantly higher than for full or resampled networks (Supplementary Material, Fig. S5), for all simulated extinction scenarios (pairwise  $t$ -test;  $t = 152.22$ ,  $121.83$ ,  $132.14$ ,  $161.45$  and  $155.72$  for  $R_g$  Trend,  $R_g N$ ,  $R_g F$ ,  $R_g R$  and  $R_I$ , respectively; d.f. = 2750 and  $P < 0.001$  in all cases). Crop–pollinator species were among the most widely occurring species in the database (median occurrence for crop–pollinators = 60% of hectads, for all bees = 30%, for all pollinators = 43%).

### DISCUSSION

#### Spatial patterns of plant–pollinator networks and relationships with agricultural land cover

Our results revealed that national-scale spatial patterns were clearly evident in all network metrics. Those for pollinator and plant generality and for connectance largely reflected well-known latitudinal gradients in GB plant and invertebrate species richness (e.g. Woodcock *et al.* 2014). This is



**Figure 4** Spatial patterns in robustness to global simulated extinctions ( $R_g$ ) as measured by area under extinction curve. Panels a, c, e, g and i show  $R_g$  with extinctions ordered by historic plant occurrence trend ( $R_g$  Trend), fertility tolerance ( $R_g N$ ), drought tolerance ( $R_g F$ ), globally randomised extinctions ( $R_g R$ ) and locally randomised extinctions ( $R_I$ ) respectively. Panels b, d, f, h and j show residuals from linear mixed models of  $R_g$  Trend,  $R_g N$ ,  $R_g F$ ,  $R_g R$  and  $R_I$ , respectively, against species richness and environmental zone. Grey-shaded cells indicate environmental zones with < 30 cells excluded from mixed models. For all panels, darker colours indicate higher values, with a linear colour stretch between maximum and minimum values.



**Table 1** Results from linear mixed models of network metrics per hectad vs. proportion of agricultural land and random effect of environmental zone for potential networks at hectad ( $10 \times 10$  km) scale across Great Britain. Results are standardised slope coefficients ( $\pm$ SE) for linear and quadratic terms (the latter only retained where significant) with level of significance (\*\*\*  $< 0.001$ , \*\*  $< 0.01$ , \*  $< 0.05$ ) and likelihood-ratio tests against a model consisting only of the random effect, and  $P$  value derived from comparing model results against 1000 randomisations. Model results are shown with and without species richness as a covariate

|  |                       | Species richness     |      | Agricultural cover<br>(linear term) |      | Agricultural cover<br>(quadratic term) |      | Likelihood-ratio<br>test |                |      | Randomisation<br>test |      |
|--|-----------------------|----------------------|------|-------------------------------------|------|--|------|--------------------------|----------------|------|-----------------------|------|
|  |                       | Slope $\pm$ SE       | Sig. | Slope $\pm$ SE                      | Sig. | Slope $\pm$ SE                         | Sig. | $\chi^2$                 | <i>P</i> value | Sig. | <i>P</i> value        | Sig. |
| Metrics  |                       |                      |      |                                     |      |  |      |                          |                |      |                       |      |
| Network species richness                                     | Total richness        |                      |      | 0.06 ( $\pm 0.12$ )                 |      |  |      | 2.22                     | 0.14           |      | 0.50                  |      |
|  | Pollinator richness   |                      |      | −0.08 ( $\pm 0.11$ )                |      |  |      | 2.01                     | 0.16           |      | 0.09                  |      |
|  | Plant richness        |                      |      | 0.15 ( $\pm 0.12$ )                 |      |  |      | 0.78                     | 0.38           |      | 0.23                  |      |
| Plant extinction scenario predictors per hectad              | Mean historic trend   |                      |      | 0.37 ( $\pm 0.07$ )                 | ***  | −0.12 ( $\pm 0.02$ )                   | ***  | 29.33                    | < 0.001        | ***  | < 0.001               | ***  |
|  | SD historic trend     |                      |      | 0.14 ( $\pm 0.12$ )                 |      |  |      | 1.00                     | 0.32           |      | 0.30                  |      |
|  | Mean Ellenberg N      |                      |      | 1.33 ( $\pm 0.53$ )                 | *    | −0.1 ( $\pm 0.02$ )                    | ***  | 14.82                    | < 0.001        | ***  | 0.03                  | *    |
|  | SD Ellenberg N        |                      |      | −0.54 ( $\pm 0.08$ )                | ***  | −0.22 ( $\pm 0.04$ )                   | ***  | 33.21                    | < 0.001        | ***  | < 0.001               | ***  |
|  | Mean Ellenberg F      |                      |      | −0.23 ( $\pm 0.07$ )                | **   |  |      | 5.07                     | 0.02           | *    | 0.01                  | **   |
|  | SD Ellenberg F        |                      |      | −0.05 ( $\pm 0.11$ )                |      |  |      | 2.56                     | 0.11           |      | 0.18                  |      |
| Network metrics (species richness not included as covariate) | Connectance           |                      |      | 0.01 ( $\pm 0.14$ )                 |      |  |      | 2.14                     | 0.14           |      | 0.27                  |      |
|  | Pollinator generality |                      |      | 0.33 ( $\pm 0.09$ )                 | ***  | −0.18 ( $\pm 0.02$ )                   | ***  | 52.29                    | < 0.001        | ***  | 0.00                  | **   |
|  | Plant generality      |                      |      | 0.08 ( $\pm 0.11$ )                 |      |  |      | 2.06                     | 0.15           |      | 0.38                  |      |
|  | $R_g$ Trend           |                      |      | 0.38 ( $\pm 0.06$ )                 | ***  | −0.09 ( $\pm 0.02$ )                   | ***  | 19.89                    | < 0.001        | ***  | < 0.001               | ***  |
|  | $R_g$ N               |                      |      | 0.49 ( $\pm 0.05$ )                 | ***  | −0.14 ( $\pm 0.02$ )                   | ***  | 49.68                    | < 0.001        | ***  | < 0.001               | ***  |
|  | $R_g$ F               |                      |      | 0.36 ( $\pm 0.09$ )                 | ***  |  |      | 10.33                    | 0.00           | **   | < 0.001               | ***  |
|  | $R_g$ R               |                      |      | 0.45 ( $\pm 0.07$ )                 | ***  |  |      | 18.65                    | < 0.001        | ***  | < 0.001               | ***  |
|  | $R_l$                 |                      |      | 0.42 ( $\pm 0.07$ )                 | ***  | −0.1 ( $\pm 0.03$ )                    | **   | 20.58                    | < 0.001        | ***  | < 0.001               | ***  |
|  | Connectance           | −1.34 ( $\pm 0.02$ ) | ***  | 0.12 ( $\pm 0.05$ )                 | *    |  |      | 0.68                     | 0.41           |      | 0.13                  |      |
| Network metrics (species richness included as covariate)     | Pollinator generality | 0.53 ( $\pm 0.02$ )  | ***  | 0.34 ( $\pm 0.07$ )                 | ***  | −0.07 ( $\pm 0.02$ )                   | **   | 13.84                    | < 0.001        | ***  | < 0.001               | ***  |
|  | Plant generality      | 0.96 ( $\pm 0.01$ )  | ***  | 0.01 ( $\pm 0.03$ )                 |      |  |      | 4.90                     | 0.03           | *    | 0.32                  |      |
|  | $R_g$ Trend           | 0.21 ( $\pm 0.03$ )  | ***  | 0.38 ( $\pm 0.05$ )                 | ***  | −0.05 ( $\pm 0.03$ )                   | *    | 19.70                    | < 0.001        | ***  | < 0.001               | ***  |
|  | $R_g$ N               | −0.12 ( $\pm 0.03$ ) | ***  | 0.49 ( $\pm 0.05$ )                 | ***  | −0.16 ( $\pm 0.02$ )                   | ***  | 54.18                    | < 0.001        | ***  | < 0.001               | ***  |
|  | $R_g$ F               | −0.12 ( $\pm 0.03$ ) | ***  | 0.36 ( $\pm 0.1$ )                  | ***  |  |      | 8.62                     | 0.00           | **   | < 0.001               | ***  |
|  | $R_g$ R               | −0.68 ( $\pm 0.04$ ) | ***  | 0.45 ( $\pm 0.09$ )                 | ***  | −0.17 ( $\pm 0.03$ )                   | ***  | 24.10                    | < 0.001        | ***  | < 0.001               | ***  |
|  | $R_l$                 | −0.57 ( $\pm 0.04$ ) | ***  | 0.42 ( $\pm 0.08$ )                 | ***  | −0.22 ( $\pm 0.03$ )                   | ***  | 42.18                    | < 0.001        | ***  | < 0.001               | ***  |
|  | Connectance           |                      |      |                                     |      |  |      |                          |                |      |                       |      |
|  | Pollinator generality |                      |      |                                     |      |  |      |                          |                |      |                       |      |

unsurprising as these metrics are a function of the connectedness between the two levels of a network and thus fundamentally affected by network species richness (Jordano 1987; Olesen & Jordano 2002; Thébault & Fontaine 2010).

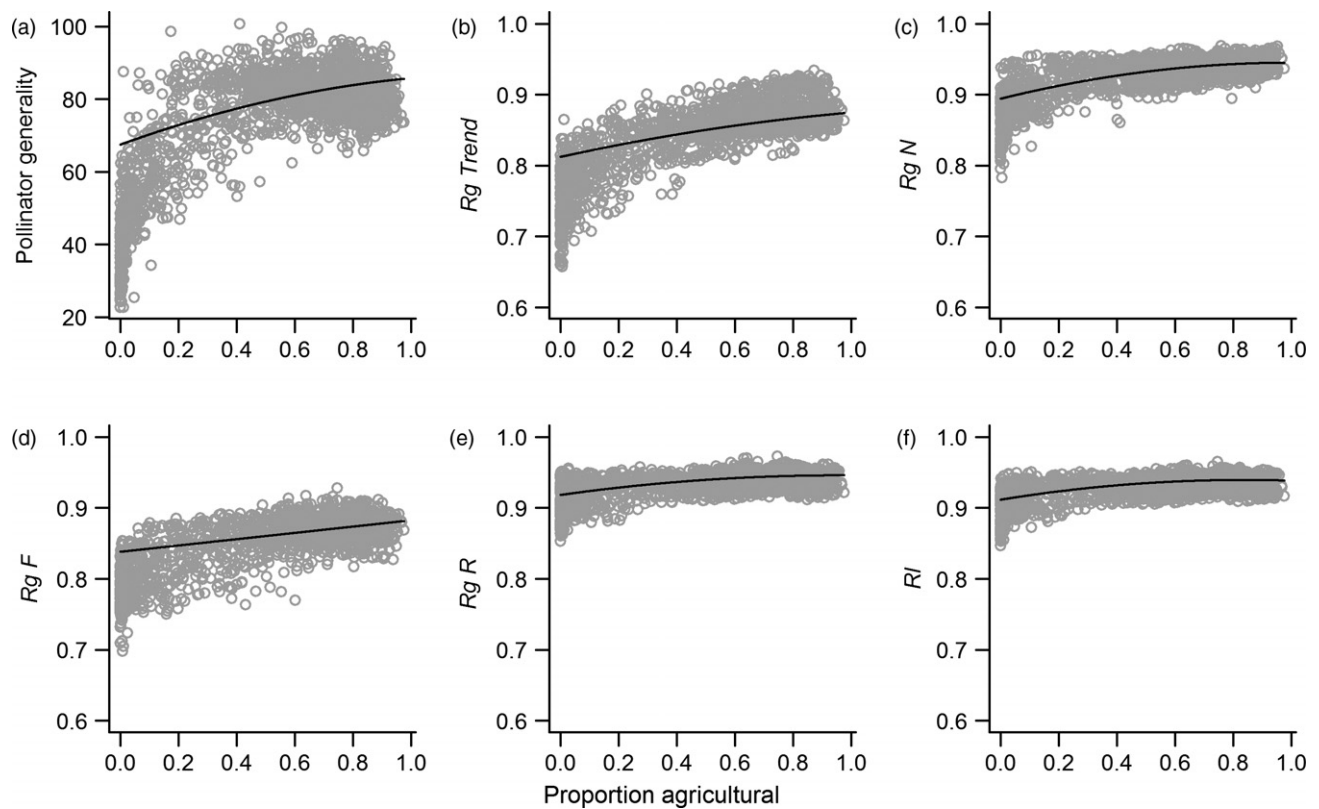
Of greater interest in terms of implications for network stability were the more complex spatial patterns of our metrics of robustness to global simulated extinctions and the positive relationship with coverage of agricultural land evident under all our extinction scenarios. This positive relationship may, at first sight, seem surprising. Highly agricultural landscapes are often considered to have depauperate plant and pollinator communities (Kremen *et al.* 2002; Potts *et al.* 2010; Ollerton *et al.* 2014). As noted by Kleijn *et al.* (2015), this is often used as justification for pollinator conservation efforts under the assumption that continued crop pollination depends upon a diverse pollinator community. While our results cannot directly shed light on the provision of pollination services, it is clear that plant–pollinator networks in landscapes with relatively high agricultural cover can exhibit higher robustness to extinction scenarios.

#### Explaining higher robustness of plant–pollinator networks in agricultural landscapes

Although it is somewhat counterintuitive that increased levels of anthropogenic disturbance (coverage of agriculture here)

can lead to increased resilience to future perturbations (as estimated by our robustness metrics), similar relationships have been observed in other ecosystems (e.g. coral reefs, Côté & Darling 2010). This might be due to positive correlations between traits that confer tolerance to past and future disturbance (Vinebrooke *et al.* 2004). Exposure to previous stressors therefore acts as a filter either extirpating vulnerable species or favouring resistant ones to produce a community more resilient to future stress. Our results suggest that the positive relationships between agricultural coverage and robustness may arise in this way from two interacting properties of the plant–pollinator networks.

First, we showed greater robustness to extinctions with increasing agricultural coverage even when extinctions were at random and irrespective of the relative vulnerabilities of the plant community to our trait- and trend-based extinction scenarios. This may be largely driven by the higher generality of pollinator communities in agricultural landscapes, thus being less reliant on individual plant species. Highly agricultural landscapes, where resources are spatially and temporally clustered and where travel between patches of resource is costly, favour the persistence of generalist pollinators (Waser *et al.* 1996). Conversely, there was no significant effect of agricultural coverage on plant generality, so the loss of each plant is no more likely to remove resources for multiple pollinators. In combination, increased pollinator generality and consistent



**Figure 5** Relationships between proportion of agricultural land cover for six network properties: a) pollinator generality b) robustness to simulated global extinctions ordered by historic plant trend ( $R_g \text{ Trend}$ ), c) robustness to simulated global extinctions ordered by Ellenberg N ( $R_g \text{ N}$ ), d) robustness to simulated global extinctions ordered by Ellenberg F ( $R_g \text{ F}$ ), e) robustness to simulated global extinctions ordered at random ( $R_g \text{ R}$ ) and f) robustness to simulated local extinctions ordered at random ( $R_l$ ). All relationships are statistically significant (see Table 1). Slopes were back transformed from the full model (i.e. with agricultural land cover, species richness and environmental zone) and show the effect of agricultural land coverage on response variables once the effects of species richness and environmental zone are accounted for.

plant generality insulate these networks against simulated extinctions.

Second, in landscapes with higher coverage of agriculture, plant communities are more liable to have already lost their most vulnerable plant species and gained more tolerant ones (evidenced by significant relationships between coverage of agriculture and mean historic trend in plant occurrence and Ellenberg values), such that global extinctions are less likely to have local impacts. Many of the severest historic declines in GB plant species are associated with agricultural expansion and intensification (Fuller 1987; Walker & Preston 2006). Therefore, many plants showing strong historic declines have already disappeared from highly agricultural areas, being replaced by species which can persist in such landscapes and with stable or increasing historic trends (Carvalho *et al.* 2013), contributing to higher robustness to extinctions ordered by plant trend. Likewise, amongst the major, lasting impacts of modern agriculture are increases in soil fertility, so agricultural hectads would have undergone replacement of those plants with low fertility tolerance (Marrs 1993; Walker & Preston 2006; Walker *et al.* 2009; Redhead *et al.* 2014), contributing to higher robustness to extinctions ordered by fertility tolerance. Furthermore, plant communities in hectads with a higher coverage of agricultural land also showed a

lower standard deviation in Ellenberg N, suggesting a homogenisation of fertility tolerances in agricultural landscapes.

Under extreme circumstances, where networks have completely extinction-prone plant communities or completely resistant ones, differences in robustness to global simulated extinctions might be driven by the second effect alone, regardless of network structure. However, this is unlikely in our data given the significant relationship with generality and robustness under randomised extinctions. Also, all hectads possessed plant communities with varying positions in the extinction sequences. For example, Figs. 2a and 2c show extinction curves for the two hectads which were, respectively, most and least robust to extinctions ordered by plant trend. From the distribution of the tick marks denoting plant extinctions from the hectad on the horizontal axes it is clear that, while these two hectads have plant communities consisting of species with differing positions in the extinction sequence, neither hectad has all its plant species at either extreme.

Of course, the observed tendency of agricultural networks to require extreme plant extinction scenarios to collapse pollinator network structure does not mean that agriculture is without detrimental effects. Simple network metrics are insufficient to capture the myriad aspects of ecological stability

(Grimm & Wissel 1997). While the networks of agricultural landscapes may be more robust to the scenarios we examined, they may also have lower levels of functional diversity. Potentially, they may also have lower functional resilience due to a homogenisation of species traits in response to the selective pressures of intensive agriculture (Woodcock *et al.* 2014; Oliver *et al.* 2015; Kaiser-Bunbury *et al.* 2017), as seen in our results for plant Ellenberg *N*.

In reality, extinctions are unlikely to proceed in a rigid linear sequence according to a single predictor. Extinction cascades (Vanbergen *et al.* 2017), rewiring (Thierry *et al.* 2011; Ramos-Jiliberto *et al.* 2012), climate change (Chen *et al.* 2011), disease (Smith *et al.* 2006) or invasive species (Bartomeus *et al.* 2008) can alter the stability of networks in unpredictable ways. However, our approach for calculating robustness to global simulated extinctions is sufficiently flexible that, where information on such effects exists, these could be incorporated into the extinction sequences.

### Crop–pollinator network properties

Our results showed that crop–pollinator networks are significantly more robust to simulated extinction scenarios than the overall networks of which they are a subset. This is probably due to the observed ubiquity and high generality of crop–pollinator species. These characteristics might be expected, as GB crop–pollinators are by definition those species pre-adapted to exploit the resource of non-native agricultural crop species growing in highly modified landscapes. Our results support the contention of Kleijn *et al.* (2015) that strategies and initiatives based on conserving crop–pollinators will provide insufficient protection for wild pollinator communities overall. More generally, our results suggest caution where such functionally specific taxa are studied in isolation of the wider communities of which they are often only a small fraction. Obviously, crop–pollinators can be threatened by a wide variety of factors other than loss of nectar sources (Vanbergen *et al.* 2013; Gill *et al.* 2016; Potts *et al.* 2016). For example, preferential loss of crop–pollinators could be triggered if association with crops results in detrimental exposure to pesticides (Stanley *et al.* 2015; Woodcock *et al.* 2016).

### Limitations of the potential network approach

Constructing potential networks from biological records has a variety of limitations and assumptions that constrain their biological realism (hence ‘potential’ networks) and affect the uncertainty of results. Perhaps the most obvious limiting factor in our networks are the biological records from which they are constructed. In particular, our data are affected by shortfalls in our knowledge of species occurrence and of their interactions (Hortal *et al.* 2015).

Regarding occurrence, although FRESCALO accounts for variation in recorder effort, there are likely to be remaining inaccuracies for rare or under-recorded species, while conversion of FRESCALO’s probabilistic outputs to binary presence/absence values may also introduce errors, particularly at species’ range boundaries. Regarding interactions, we know that our coverage of GB plants, pollinators and the

interactions between them are incomplete (see Appendix S2). Our potential networks may exhibit either ‘missing’ or ‘forbidden’ links in some hectads (Olesen *et al.* 2010) as they do not account for variation in interactions due to flower phenology (Basilio *et al.* 2006; Rafferty & Ives 2011), pollinator life-history (Vieira & Almeida-Neto 2015; Vanbergen *et al.* 2017) or pollinator resource-switching (Thomsen *et al.* 2017).

Our exploration of some of these sources of uncertainty (see Appendices S4 and S5) suggests that uncertainties arising from occurrence and/or interaction data affect hectad-level networks in ways that are relatively consistent across space. While both sources of uncertainty affect the accuracy of individual potential networks they are far less likely to introduce a systemic bias which would affect our observed spatial patterns and relationships with agricultural land. Therefore, despite these limitations, we suggest that our potential networks properties and the spatial patterns we observe are broadly representative of real-world networks (see Appendix S5).

### CONCLUSIONS

Our results demonstrate the ability of potential networks constructed from biological records to provide new insights into spatial patterns of ecological networks across national scales that would be impossible to monitor using conventional direct observation approaches. The positive relationship between agricultural cover and robustness to a range of extinction scenarios supports previous observations that anthropogenic disturbance can result in ecological networks which are more robust to further perturbation. Furthermore, from our results, crop–pollinator networks are not representative of wider plant–pollinator networks, such that targeting landscape management for the retention of crop pollination may be entirely insufficient to conserve wider biodiversity (Kleijn *et al.* 2015).

Our findings suggest potentially productive fields of further investigation, including further investigation of the mechanisms underpinning spatial patterns in network properties, validation of potential networks against those constructed from large-scale molecular data and exploration of more complex scenarios of extinction, invasion or restoration. In the future, the production of potential networks from biological records is likely to become easier and more accurate, as new technology and methods increase the quality and quantity of biological records (Tulloch *et al.* 2013; Gray *et al.* 2014; August *et al.* 2015a; Powney & Isaac 2015) and novel molecular techniques increase the potential for wide-scale validation (Keller *et al.* 2015; Richardson *et al.* 2015; Bohan *et al.* 2017; Pornon *et al.* 2017).

### ACKNOWLEDGEMENTS

We thank the committee and members of the Bees, Wasps and Ants Recording Society, Hoverfly Recording Scheme, Butterfly Conservation and Botanical Society of Britain & Ireland for making available the biological records. We also thank the many hundreds of volunteers who have contributed to the biological recording schemes. We thank Hannah Dean, Stephanie Rorke, Charlotte Coombes and Rob Dyer for

assistance with compilation of the plant–pollinator interactions database; Oli Pescott and Tom Humphries for assistance with BSBI data; Gary Powney and Tom August for guidance on the use of FRESALO in R; and Mike Edwards (BWARS) for advice on plant–pollinator interactions. We also extend our thanks to four reviewers who provided insightful and constructive comments on previous versions of this paper. This work was supported by NERC national capability funding to CEH and the Natural Environment Research Council (NERC) and the Biotechnology and Biological Sciences Research Council (BBSRC) under research programme NE/N018125/1 LTS-M ASSIST – Achieving Sustainable Agricultural Systems ([www.assist.ceh.ac.uk](http://www.assist.ceh.ac.uk)).

## AUTHORSHIP

JWR performed the data handling, conducted spatial and statistical analyses, wrote the first draft of the manuscript and undertook revisions. THO and AJV coordinated the project under which this work was performed. BAW and MJOP supported ecological network analyses. RFP conceived and coordinated the compilation of plant–pollinator interaction records. All authors contributed to writing of the manuscript.

## DATA ACCESSIBILITY

Plant–pollinator network data and metadata are available from the NERC Environmental Information Data Centre (EIDC) at [doi.org/10.5285/6d8d5cb5-bd54-4da7-903a-15bd4bbd531b](https://doi.org/10.5285/6d8d5cb5-bd54-4da7-903a-15bd4bbd531b). Biological records are available on request from the relevant recording schemes.

## REFERENCES

- Asher, J. (1997). *Butterflies for the New Millennium - Project and Programme*. ITE Monks Wood, Huntingdon.
- August, T., Harvey, M., Lightfoot, P., Kilbey, D., Papadopoulos, T. & Jepson, P. (2015a). Emerging technologies for biological recording. *Biol. J. Lin. Soc.*, 115, 731–749.
- August, T., Powney, G., Harrower, C., Hill, M. & Isaac, N.J.B. (2015b). sparta: Trend analysis for unstructured data. R package version 0.1.30.
- Ballantyne, G., Baldock, K.C.R. & Willmer, P.G. (2015). Constructing more informative plant–pollinator networks: visitation and pollen deposition networks in a heathland plant community. *Proc. Biol. Sci.*, 282, 20151130.
- Bartomeus, I., Vilà, M. & Santamaría, L. (2008). Contrasting effects of invasive plants in plant–pollinator networks. *Oecologia*, 155, 761–770.
- Basilio, A.M., Medan, D., Torretta, J.P. & Bartoloni, N.J. (2006). A year-long plant–pollinator network. *Austral Ecol.*, 31, 975–983.
- Bersier, L.-F., Banašek-Richter, C. & Cattin, M.-F. (2002). Quantitative descriptors of food-web matrices. *Ecology*, 83, 2394–2407.
- Bishop, T.R., Botham, M.S., Fox, R., Leather, S.R., Chapman, D.S. & Oliver, T.H. (2013). The utility of distribution data in predicting phenology. *Methods Ecol. Evol.*, 4, 1024–1032.
- Bohan, D.A., Vacher, C., Tamaddoni-Nezhad, A., Raybould, A., Dumbrell, A.J. & Woodward, G. (2017). Next-generation global biomonitoring: large-scale, automated reconstruction of ecological networks. *Trends Ecol. Evol.*, 32, 477–487.
- Bunce, R.G.H., Barr, C.J., Clarke, R.T., Howard, D.C. & Scott, W.A. (2007). ITE land classification of Great Britain 2007. NERC Environmental Information Data Centre, <https://doi.org/10.5285/5f0605e4-aa2a-48ab-b47c-bf5510823e8f>.
- Burgos, E., Ceva, H., Perazzo, R.P.J., Devoto, M., Medan, D., Zimmermann, M. *et al.* (2007). Why nestedness in mutualistic networks? *J. Theor. Biol.*, 249, 307–313.
- Burkle, L.A. & Alarcón, R. (2011). The future of plant–pollinator diversity: understanding interaction networks across time, space, and global change. *Am. J. Bot.*, 98, 528–538.
- Carstensen, D.W., Sabatino, M., Trøjelsgaard, K. & Morellato, L.P.C. (2014). Beta diversity of plant–pollinator networks and the spatial turnover of pairwise interactions. *PLoS ONE*, 9, e112903.
- Carvalho, L.G., Kunin, W.E., Keil, P., Aguirre-Gutiérrez, J., Ellis, W.N., Fox, R. *et al.* (2013). Species richness declines and biotic homogenisation have slowed down for NW-European pollinators and plants. *Ecol. Lett.*, 16, 870–878.
- Carvell, C. (2002). Habitat use and conservation of bumblebees (*Bombus* spp.) under different grassland management regimes. *Biol. Cons.*, 103, 33–49.
- Chacoff, N.P., Vázquez, D.P., Lomáscolo, S.B., Stevani, E.L., Dorado, J. & Padrón, B. (2012). Evaluating sampling completeness in a desert plant–pollinator network. *J. Anim. Ecol.*, 81, 190–200.
- Chen, I.-C., Hill, J.K., Ohlemüller, R., Roy, D.B. & Thomas, C.D. (2011). Rapid range shifts of species associated with high levels of climate warming. *Science*, 333, 1024–1026.
- Côté, I.M. & Darling, E.S. (2010). Rethinking ecosystem resilience in the face of climate change. *PLoS Biol.*, 8, e1000438.
- Dormann, C.F., Fründ, J., Blüthgen, N. & Gruber, B. (2009). Indices, graphs and null models: analyzing bipartite ecological networks. *Open Ecol. J.*, 2, 7–24.
- Dunne, J.A., Williams, R.J. & Martinez, N.D. (2002). Food-web structure and network theory: the role of connectance and size. *Proc. Natl Acad. Sci.*, 99, 12917–12922.
- Dyer, R.J., Gillings, S., Pywell, R.F., Fox, R., Roy, D.B. & Oliver, T.H. (2016). Developing a biodiversity-based indicator for large-scale environmental assessment: a case study of proposed shale gas extraction sites in Britain. *J. Appl. Ecol.*, 54, 872–882.
- Eilers, E.J., Kremen, C., Greenleaf, S.S., Garber, A.K. & Klein, A.-M. (2011). Contribution of pollinator-mediated crops to nutrients in the human food supply. *PLoS ONE*, 6, e21363.
- Falcão, J.C.F., Dáttilo, W. & Rico-Gray, V. (2016). Sampling effort differences can lead to biased conclusions on the architecture of ant–plant interaction networks. *Ecol. Complex.*, 25, 44–52.
- Falk, S.J. (2015). *Field Guide to the Bees of Great Britain and Ireland*. British Wildlife Publishing, Oxford, UK.
- Fitter, A.H. & Peat, H.J. (1994). The ecological flora database. *J. Ecol.*, 82, 415–425.
- Fortin, M.-J. & Jaquez, G.M. (2000). Randomization tests and spatially auto-correlated data. *Bull. Ecol. Soc. Am.*, 81, 201–205.
- Forup, M.L., Henson, K.S.E., Craze, P.G. & Memmott, J. (2008). The restoration of ecological interactions: plant–pollinator networks on ancient and restored heathlands. *J. Appl. Ecol.*, 45, 742–752.
- Fox, R., Oliver, T.H., Harrower, C., Parsons, M.S., Thomas, C.D. & Roy, D.B. (2014). Long-term changes to the frequency of occurrence of British moths are consistent with opposing and synergistic effects of climate and land-use changes. *J. Appl. Ecol.*, 51, 949–957.
- Free, J.B. (1993). *Insect Pollination of Crops*. Academic press, Cambridge, MA.
- Fuller, R.M. (1987). The changing extent and conservation interest of lowland grasslands in England and Wales: a review of grassland surveys 1930–1984. *Biol. Cons.*, 40, 281–300.
- Garratt, M.P.D., Coston, D.J., Truslove, C.L., Lappage, M.G., Polce, C., Dean, R. *et al.* (2014). The identity of crop pollinators helps target conservation for improved ecosystem services. *Biol. Cons.*, 169, 128–135.
- Gill, R.J., Baldock, K.C.R., Brown, M.J.F., Cresswell, J.E., Dicks, L.V., Fountain, M.T. *et al.* (2016). Protecting an ecosystem service: approaches to understanding and mitigating threats to wild insect



- pollinators. In *Advances in Ecological Research*. (eds Guy, W. & David, A.B.). Academic Press, Cambridge, MA, pp. 135–206.
- Gray, C., Baird, D.J., Baumgartner, S., Jacob, U., Jenkins, G.B., O’Gorman, E.J. *et al.* (2014). FORUM: ecological networks: the missing links in biomonitoring science. *J. Appl. Ecol.*, 51, 1444–1449.
- Grimm, V. & Wissel, C. (1997). Babel, or the ecological stability discussions: an inventory and analysis of terminology and a guide for avoiding confusion. *Oecologia*, 109, 323–334.
- Hill, M.O. (2012). Local frequency as a key to interpreting species occurrence data when recording effort is not known. *Methods Ecol. Evol.*, 3, 195–205.
- Hill, M.O., Preston, C.D. & Roy, D. (2004). *PLANTATT-Attributes of British and Irish Plants: Status, Size, Life History, Geography and Habitats*. Centre for Ecology & Hydrology, Lancaster, UK.
- Hortal, J., de Bello, F., Diniz-Filho, J.A.F., Lewinsohn, T.M., Lobo, J.M. & Ladle, R.J. (2015). Seven shortfalls that beset large-scale knowledge of biodiversity. *Annu. Rev. Ecol. Evol. Syst.*, 46, 523–549.
- Isaac, N.J.B., van Strien, A.J., August, T.A., de Zeeuw, M.P. & Roy, D.B. (2014). Statistics for citizen science: extracting signals of change from noisy ecological data. *Methods Ecol. Evol.*, 5, 1052–1060.
- Jordano, P. (1987). Patterns of mutualistic interactions in pollination and seed dispersal: connectance, dependence asymmetries, and coevolution. *Am. Nat.*, 000, 657–677.
- Kaartinen, R. & Roslin, T. (2012). High temporal consistency in quantitative food web structure in the face of extreme species turnover. *Oikos*, 121, 1771–1782.
- Kaiser-Bunbury, C.N., Muff, S., Memmott, J., Müller, C.B. & Caflich, A. (2010). The robustness of pollination networks to the loss of species and interactions: a quantitative approach incorporating pollinator behaviour. *Ecol. Lett.*, 13, 442–452.
- Kaiser-Bunbury, C.N., Mougal, J., Whittington, A.E., Valentin, T., Gabriel, R., Olesen, J.M. *et al.* (2017). Ecosystem restoration strengthens pollination network resilience and function. *Nature*, 542, 223–227.
- Keller, A., Danner, N., Grimmer, G., Ankenbrand, M., Ohe, K., Ohe, W. *et al.* (2015). Evaluating multiplexed next-generation sequencing as a method in palynology for mixed pollen samples. *Plant Biol.*, 17, 558–566.
- Kemp, J.E., Evans, D.M., Augustyn, W.J. & Ellis, A.G. (2017). Invariant antagonistic network structure despite high spatial and temporal turnover of interactions. *Ecography*, 40, 1315–1324.
- Kleijn, D., Winfree, R., Bartomeus, I., Carvalheiro, L.G., Henry, M., Isaacs, R. *et al.* (2015). Delivery of crop pollination services is an insufficient argument for wild pollinator conservation. *Nat. Commun.*, 6, 7414.
- Kluser, S. & Peduzzi, P. (2007). Global pollinator decline: a literature review. UNEP/GRID-Europe, Geneva.
- Kremen, C., Williams, N.M. & Thorp, R.W. (2002). Crop pollination from native bees at risk from agricultural intensification. *Proc. Natl Acad. Sci.*, 99, 16812–16816.
- Marrs, R. (1993). Soil fertility and nature conservation in Europe: theoretical considerations and practical management solutions. *Adv. Ecol. Res.*, 24, 241–300.
- Maskell, L.C., Smart, S.M., Bullock, J.M., Thompson, K. & Stevens, C.J. (2010). Nitrogen deposition causes widespread loss of species richness in British habitats. *Glob. Change Biol.*, 16, 671–679.
- Memmott, J., Waser, N.M. & Price, M.V. (2004). Tolerance of pollination networks to species extinctions. *Proc. Biol. Sci.*, 271, 2605–2611.
- Memmott, J., Craze, P.G., Waser, N.M. & Price, M.V. (2007). Global warming and the disruption of plant–pollinator interactions. *Ecol. Lett.*, 10, 710–717.
- Menz, M.H.M., Phillips, R.D., Winfree, R., Kremen, C., Aizen, M.A., Johnson, S.D. *et al.* (2011). Reconnecting plants and pollinators: challenges in the restoration of pollination mutualisms. *Trends Plant Sci.*, 16, 4–12.
- Minckley, R.L. & Roulston, T. (2006). Incidental mutualisms and pollen specialization among bees. In *Plant–Pollinator Interactions: From Specialization to Generalization*. (eds Waser, N.M. & Ollerton, J.). The University of Chicago Press, Chicago and London, pp. 69–98.
- Morales-Castilla, I., Matias, M.G., Gravel, D. & Araújo, M.B. (2015). Inferring biotic interactions from proxies. *Trends Ecol. Evol.*, 30, 347–356.
- Morris, R.K. (1998). *Hoverflies of Surrey*. Surrey Wildlife Trust, Surrey, UK.
- Morris, R.J., Gripenberg, S., Lewis, O.T. & Roslin, T. (2014). Antagonistic interaction networks are structured independently of latitude and host guild. *Ecol. Lett.*, 17, 340–349.
- Morton, D., Rowland, C., Wood, C., Meek, L., Marston, C., Smith, G. *et al.* (2011). Final report for LCM2007 - the new UK land cover map. In: NERC/Centre for Ecology and Hydrology, p. 112.
- Moruela-Holme, N., Blonder, B., Sandel, B., McGill, B.J., Peet, R.K., Ott, J.E. *et al.* (2016). A network approach for inferring species associations from co-occurrence data. *Ecography*, 39, 1139–1150.
- Olesen, J.M. & Jordano, P. (2002). Geographic patterns in plant–pollinator mutualistic networks. *Ecology*, 83, 2416–2424.
- Olesen, J.M., Bascompte, J., Dupont, Y.L., Elberling, H., Rasmussen, C. & Jordano, P. (2010). Missing and forbidden links in mutualistic networks. *Proc. Biol. Sci.*, 278, 725–732.
- Oliver, T.H., Heard, M.S., Isaac, N.J.B., Roy, D.B., Procter, D., Eigenbrod, F. *et al.* (2015). Biodiversity and resilience of ecosystem functions. *Trends Ecol. Evol.*, 30, 673–684.
- Ollerton, J., Erenler, H., Edwards, M. & Crockett, R. (2014). Extinctions of aculeate pollinators in Britain and the role of large-scale agricultural changes. *Science*, 346, 1360–1362.
- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D. & R Core Team (2015). nlme: Linear and Nonlinear Mixed Effects Models. In: R package.
- Pornon, A., Andalo, C., Burrus, M. & Escaravage, N. (2017). DNA metabarcoding data unveils invisible pollination networks. *Sci. Rep.*, 7, 16828.
- Potts, S.G., Biesmeijer, J.C., Kremen, C., Neumann, P., Schweiger, O. & Kunin, W.E. (2010). Global pollinator declines: trends, impacts and drivers. *Trends Ecol. Evol.*, 25, 345–353.
- Potts, S.G., Imperatriz-Fonseca, V., Ngo, H.T., Aizen, M.A., Biesmeijer, J.C., Breeze, T.D. *et al.* (2016). Safeguarding pollinators and their values to human well-being. *Nature*, 540, 220–229.
- Powney, G.D. & Isaac, N.J. (2015). Beyond maps: a review of the applications of biological records. *Biol. J. Lin. Soc.*, 115, 532–542.
- R Core Team (2017). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Rader, R., Bartomeus, I., Garibaldi, L.A., Garratt, M.P.D., Howlett, B.G., Winfree, R. *et al.* (2016). Non-bee insects are important contributors to global crop pollination. *Proc. Natl Acad. Sci.*, 113, 146–151.
- Rafferty, N.E. & Ives, A.R. (2011). Effects of experimental shifts in flowering phenology on plant–pollinator interactions. *Ecol. Lett.*, 14, 69–74.
- Ramos-Jiliberto, R., Valdovinos, F.S., Moisset de Espanés, P. & Flores, J.D. (2012). Topological plasticity increases robustness of mutualistic networks. *J. Anim. Ecol.*, 81, 896–904.
- Redhead, J.W., Sheail, J., Bullock, J.M., Ferreruela, A., Walker, K.J. & Pywell, R.F. (2014). The natural regeneration of calcareous grassland at a landscape scale: 150 years of plant community re-assembly on Salisbury Plain, UK. *Appl. Veg. Sci.*, 17, 408–418.
- Richardson, R.T., Lin, C.-H., Sponsler, D.B., Quijia, J.O., Goodell, K. & Johnson, R.M. (2015). Application of ITS2 metabarcoding to determine the provenance of pollen collected by honey bees in an agroecosystem. *Appl. Plant Sci.*, 3, 1400066.
- Robinson, R.A. & Sutherland, W.J. (2002). Post-war changes in arable farming and biodiversity in Great Britain. *J. Appl. Ecol.*, 39, 157–176.
- Smith, K.F., Sax, D.F. & Lafferty, K.D. (2006). Evidence for the role of infectious disease in species extinction and endangerment. *Conserv. Biol.*, 20, 1349–1357.
- Stanley, D.A., Garratt, M.P., Wickens, J.B., Wickens, V.J., Potts, S.G. & Raine, N.E. (2015). Neonicotinoid pesticide exposure impairs crop pollination services provided by bumblebees. *Nature*, 528, 548–550.

- Stevens, C.J., Dise, N.B., Gowing, D.J.G. & Mountford, J.O. (2006). Loss of forb diversity in relation to nitrogen deposition in the UK: regional trends and potential controls. *Glob. Change Biol.*, 12, 1823–1833.
- Stubbs, A.E. & Falk, S.J. (2002). *British Hoverflies: An Illustrated Identification Guide*. British Entomological and Natural History Society, Reading, UK.
- Thébault, E. & Fontaine, C. (2010). Stability of ecological communities and the architecture of mutualistic and trophic networks. *Science*, 329, 853–856.
- Thierry, A., Beckerman, A.P., Warren, P.H., Williams, R.J., Cole, A.J. & Petchey, O.L. (2011). Adaptive foraging and the rewiring of size-structured food webs following extinctions. *Basic Appl. Ecol.*, 12, 562–570.
- Thomas, J. (2010). *Butterflies of Britain and Ireland*. British Wildlife Publishing, Oxford, UK.
- Thomsen, M.S., Garcia, C., Bolam, S.G., Parker, R., Godbold, J.A. & Solan, M. (2017). Consequences of biodiversity loss diverge from expectation due to post-extinction compensatory responses. *Sci. Rep.*, 7, 43695.
- Thuiller, W., Lavorel, S., Araújo, M.B., Sykes, M.T. & Prentice, I.C. (2005). Climate change threats to plant diversity in Europe. *Proc. Natl Acad. Sci. USA*, 102, 8245–8250.
- Trøjelsgaard, K., Jordano, P., Carstensen, D.W. & Olesen, J.M. (2015). Geographical variation in mutualistic networks: similarity, turnover and partner fidelity. *Proc. Biol. Sci.*, 282, 20142925.
- Tulloch, A.I.T., Possingham, H.P., Joseph, L.N., Szabo, J. & Martin, T.G. (2013). Realising the full potential of citizen science monitoring programs. *Biol. Cons.*, 165, 128–138.
- Tylianakis, J.M., Tscharntke, T. & Lewis, O.T. (2007). Habitat modification alters the structure of tropical host–parasitoid food webs. *Nature*, 445, 202–205.
- Vanbergen, A.J., Baude, M., Biesmeijer, J.C., Britton, N.F., Brown, M.J.F., Brown, M. *et al.* (2013). Threats to an ecosystem service: pressures on pollinators. *Front. Ecol. Environ.*, 11, 251–259.
- Vanbergen, A.J., Woodcock, B.A., Heard, M.S. & Chapman, D.S. (2017). Network size, structure, and mutualism dependence affect the propensity for plant–pollinator extinction cascades. *Funct. Ecol.*, 31, 1285–1293.
- Vázquez, D.P., Blüthgen, N., Cagnolo, L. & Chacoff, N.P. (2009). Uniting pattern and process in plant–animal mutualistic networks: a review. *Ann. Bot.*, 103, 1445–1457.
- Vieira, M.C. & Almeida-Neto, M. (2015). A simple stochastic model for complex coextinctions in mutualistic networks: robustness decreases with connectance. *Ecol. Lett.*, 18, 144–152.
- Vinebrooke, R.D., Cottingham, K.L., Norberg, M.S.J., Dodson, S.I., Maberly, S.C. & Sommer, U. (2004). Impacts of multiple stressors on biodiversity and ecosystem functioning: the role of species co-tolerance. *Oikos*, 104, 451–457.
- Walker, K.J. & Preston, C.D. (2006). Ecological predictors of extinction risk in the flora of lowland England, UK. *Biodivers. Conserv.*, 15, 1913–1942.
- Walker, K.J., Preston, C.D. & Boon, C.R. (2009). Fifty years of change in an area of intensive agriculture: plant trait responses to habitat modification and conservation, Bedfordshire, England. *Biodivers. Conserv.*, 18, 3597.
- Waser, N.M., Chittka, L., Price, M.V., Williams, N.M. & Ollerton, J. (1996). Generalization in pollination systems, and why it matters. *Ecology*, 77, 1043–1060.
- Watts, G., Battarbee, R.W., Bloomfield, J.P., Crossman, J., Daccache, A., Durance, I. *et al.* (2015). Climate change and water in the UK – past changes and future prospects. *Prog. Phys. Geogr.*, 39, 6–28.
- Welti, E.A.R. & Joern, A. (2015). Structure of trophic and mutualistic networks across broad environmental gradients. *Ecol. Evol.*, 5, 326–334.
- Winfrey, R. (2008). Pollinator-dependent crops: an increasingly risky business. *Curr. Biol.*, 18, R968–R969.
- Woodcock, B., Edwards, M., Redhead, J., Meek, W., Nuttall, P., Falk, S. *et al.* (2013). Crop flower visitation by honeybees, bumblebees and solitary bees: Behavioural differences and diversity responses to landscape. *Agr. Ecosyst. Environ.*, 171, 1–8.
- Woodcock, B.A., Harrower, C., Redhead, J., Edwards, M., Vanbergen, A.J., Heard, M.S. *et al.* (2014). National patterns of functional diversity and redundancy in predatory ground beetles and bees associated with key UK arable crops. *J. Appl. Ecol.*, 51, 142–151.
- Woodcock, B.A., Isaac, N.J., Bullock, J.M., Roy, D.B., Garthwaite, D.G., Crowe, A. *et al.* (2016). Impacts of neonicotinoid use on long-term population changes in wild bees in England. *Nat. Commun.*, 7, 12459.

Editor, José Maria Gomez

Manuscript received 9 April 2018

First decision made 12 May 2018

Second decision made 3 August 2018

Manuscript accepted 8 August 2018